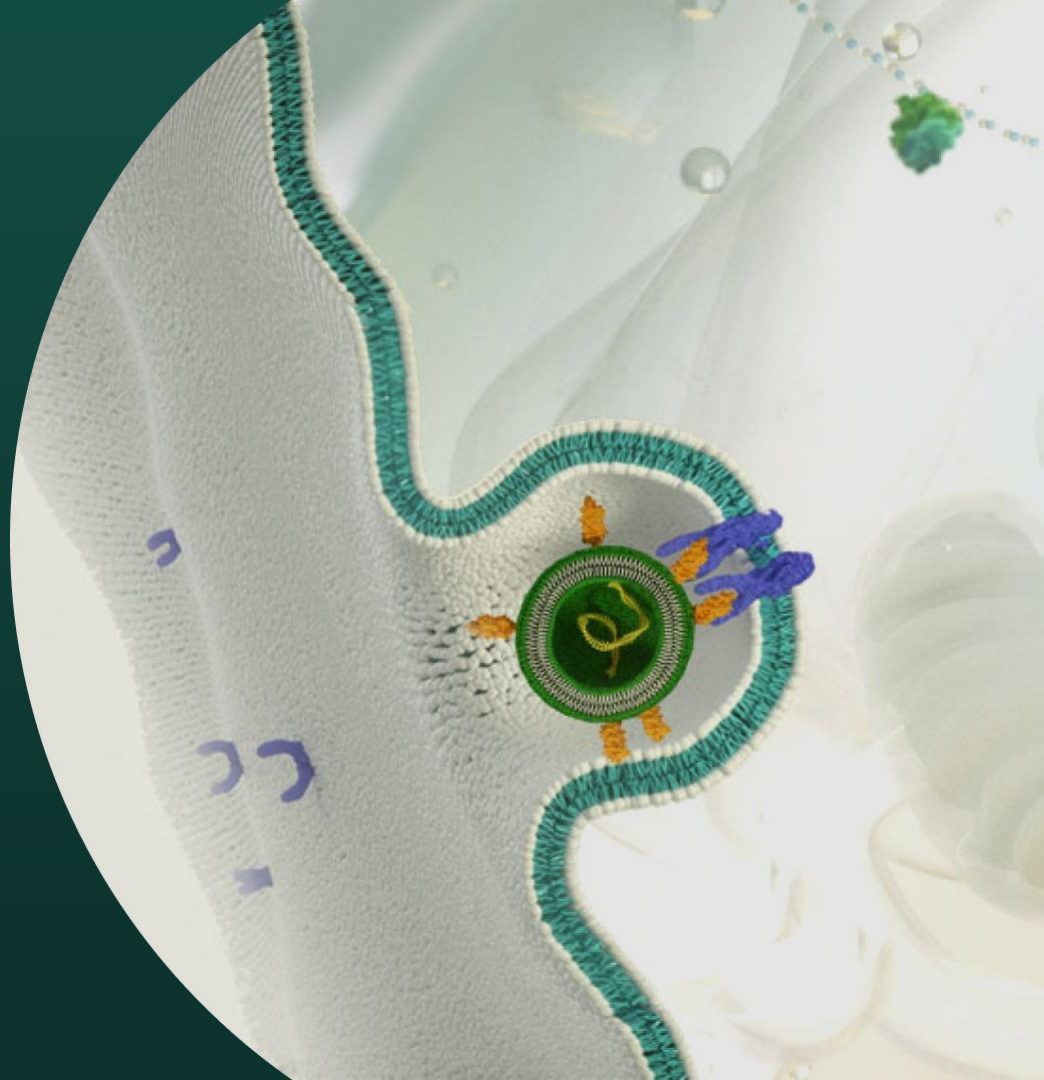


# Ionizable lipid impurities drive RNA-lipid adduct formation in LNP, impacting in vivo expression

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# Who We Are



## OUR LNP TECHNOLOGY

First Clinically Approved:

- RNA interference-based Medicine (**Onpattro®**)
- mRNA Vaccine (**Comirnaty®**)
- Personalized CRISPR gene editing therapy (**baby KJ**)



## WHO WE WORK WITH

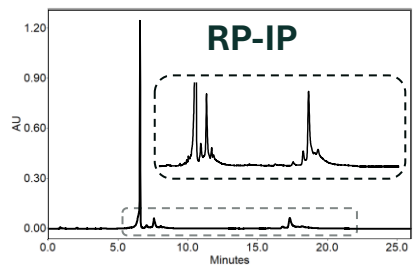
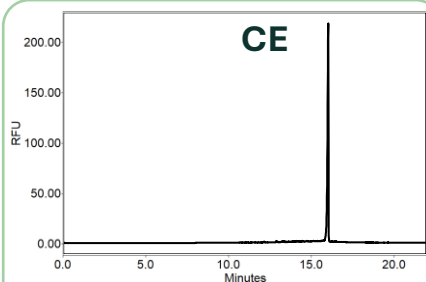
- Cutting edge **pharmaceutical & biotechnology** companies
- Leading **academics in universities & institutes**
- **Foundations & NGOs**



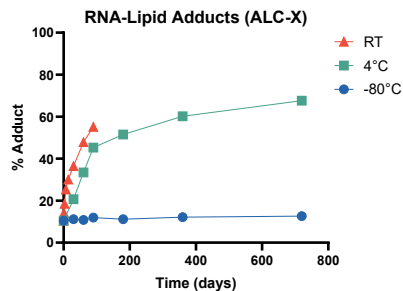
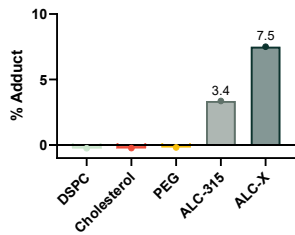
## HOW WE WORK

We exclusively work in collaboration with partners and are **focused on supporting our partners** to bring their drug products to patients

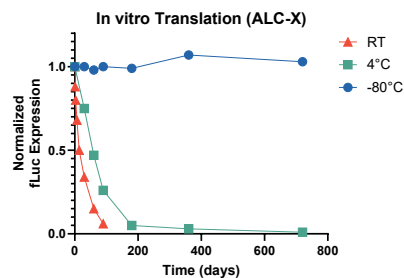
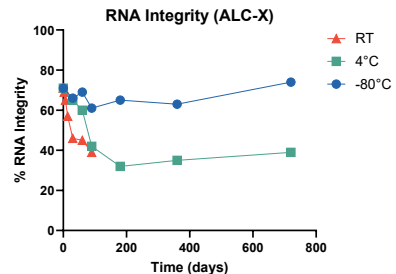
# What are RNA-Lipid Adducts?



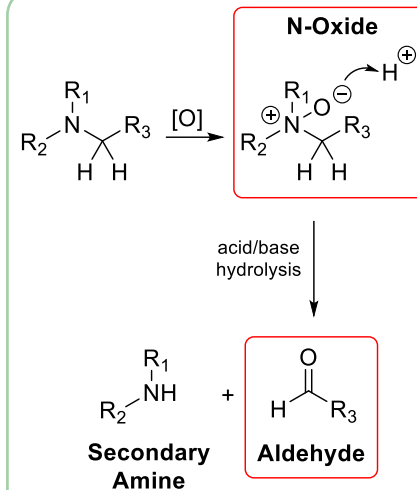
- RNA-lipid impurity undetected by typical mRNA purity techniques
- Detected as late-peaks in RP-IP



- Adduct formation linked to the ionizable lipid
- Formation is dependent on time and temperature



- RNA-lipid adducts, together with RNA integrity, **reduce in vitro protein expression**



- N-oxide forms through tertiary amine oxidation
- Acid/base-catalyzed hydrolysis generates aldehydes and secondary amines

# Investigative Goals

**01**

## Develop Analytical Methods

Implement analytical methods to measure RNA-lipid adducts and adduct forming impurities

**02**

## Impurity Testing

Screen multiple ionizable lipids and lipid lots for adduct formation and adduct forming impurities

**03**

## Determine Root Cause

Identify the main impurities contributing to RNA-lipid adduct formation

**04**

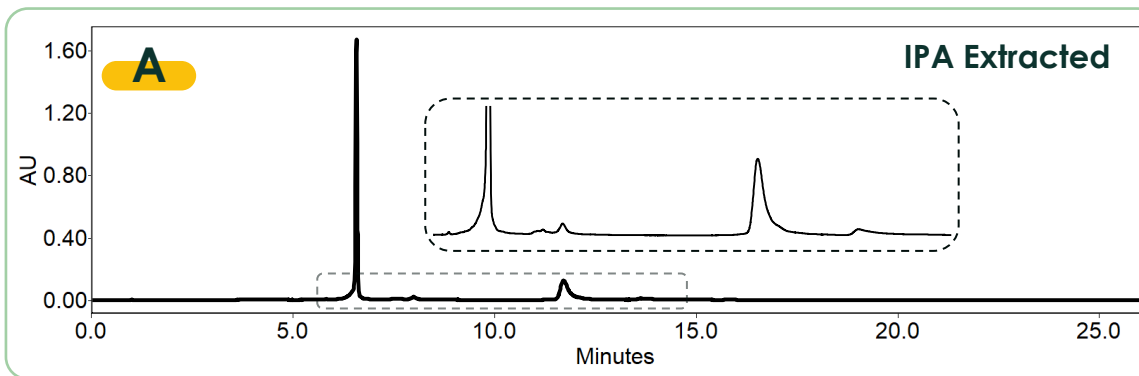
## *In vivo* Expression and Safety

Assess impact to *in vivo* expression and safety in mice with high level of impurities

# Measuring RNA-Lipid Adducts

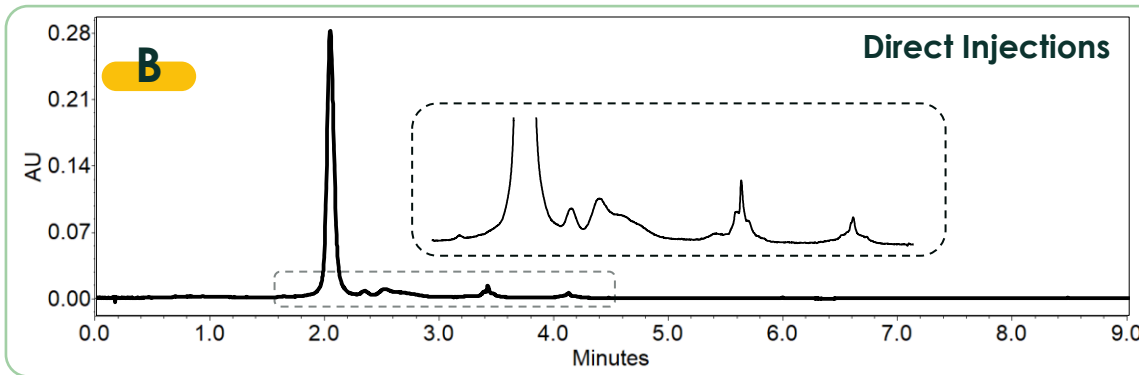
## Binary Reactions

- Mixture of ionizable lipid (ethanol) and buffered mRNA (aqueous)
- Incubate at room temperature
- Extract RNA via IPA precipitation and analysis on **Method A** or direct analysis on **Method B**



## LNP Analysis

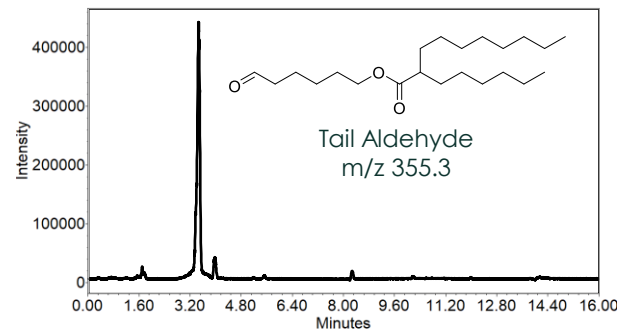
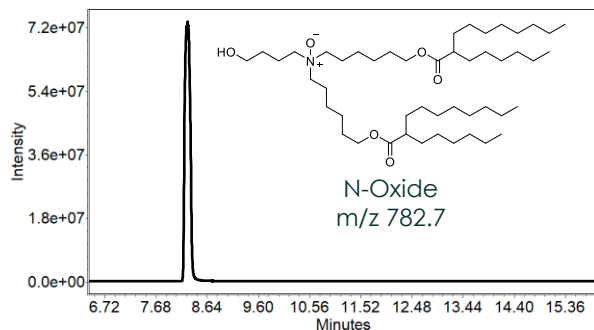
- Extract RNA via IPA precipitation and analyze on **Method A**
- Disrupt LNP with surfactant and analyze directly on **Method B**



# Measuring Lipid Impurities

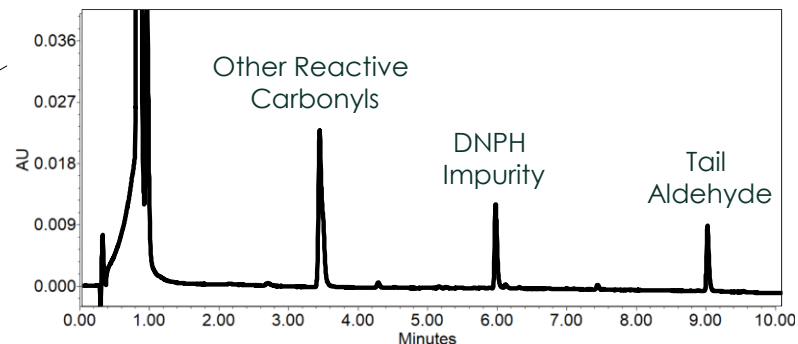
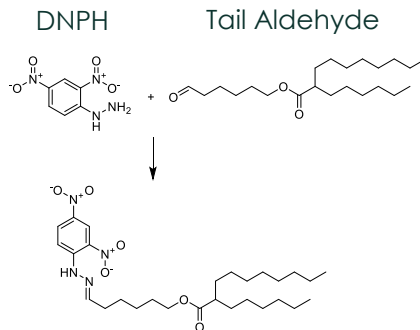
## Targeted Impurity Analysis

- Direct impurity detection in raw lipids or LNP by UPLC with MS detection
- Quantification using internally synthesized standards

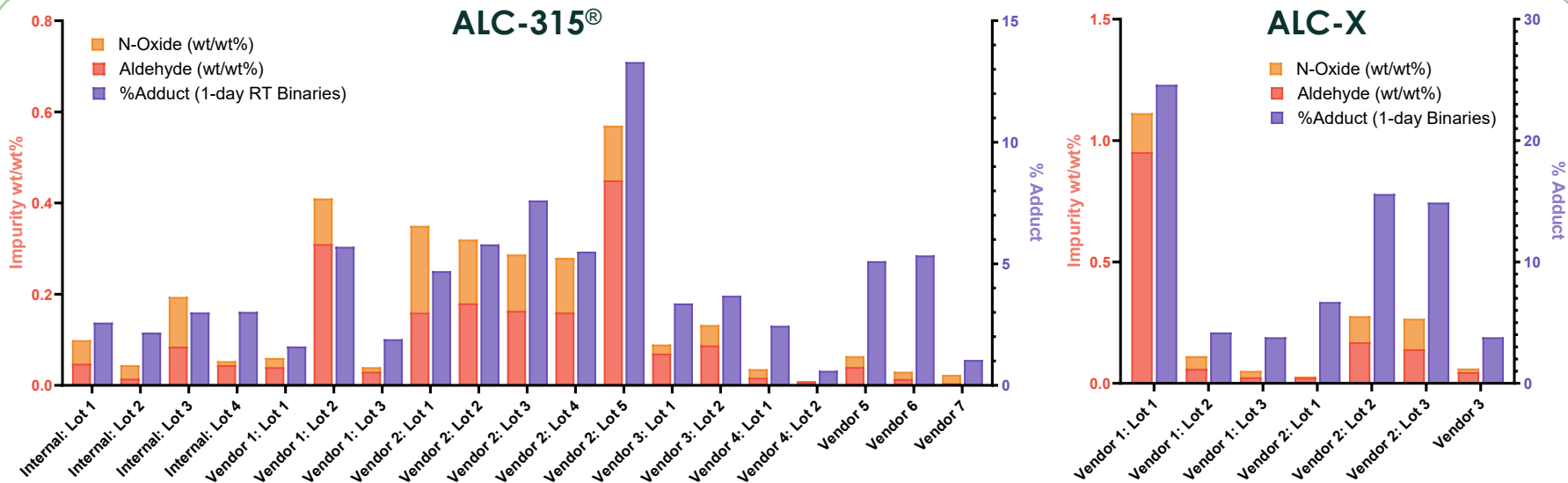


## Derivatization with DNPH

- 2,4-Dinitrophenylhydrazine (DNPH) reacts specifically with carbonyls (ketones, aldehydes) to form derivatization products
- Inject into UPLC-PDA/MS monitoring for mass and  $A_{360}$

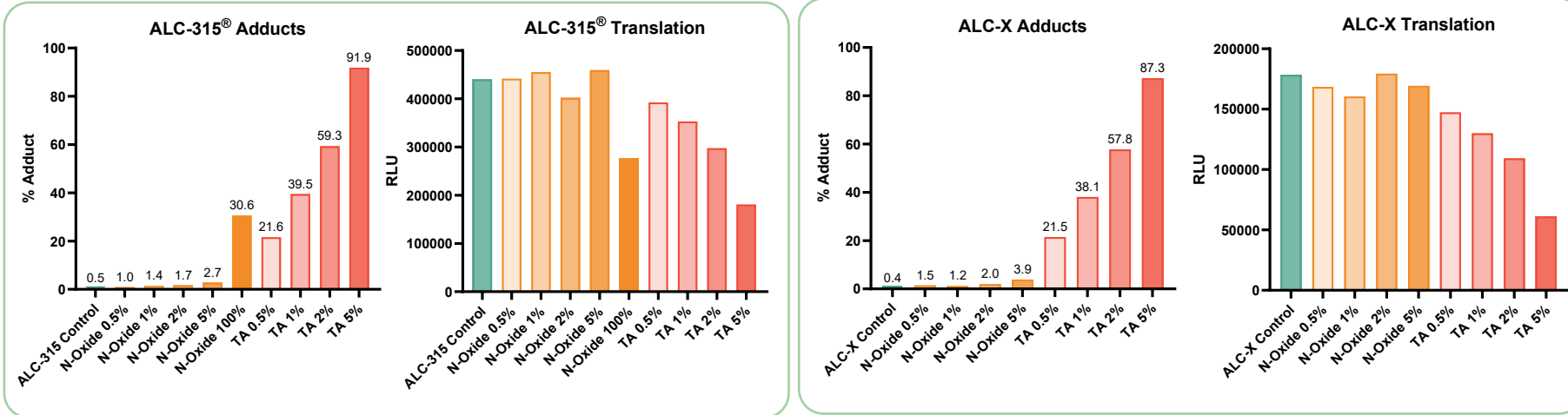


# Impurity Screening of ALC-315<sup>®</sup> and ALC-X Lots



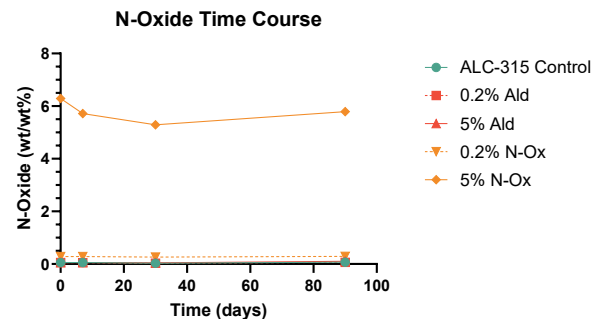
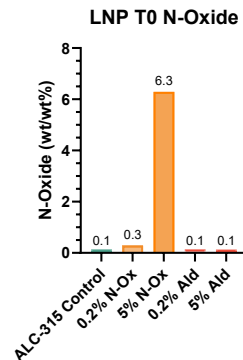
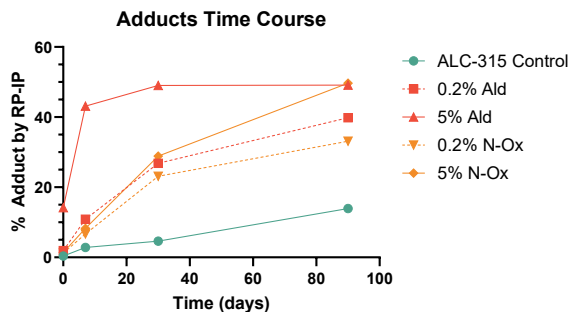
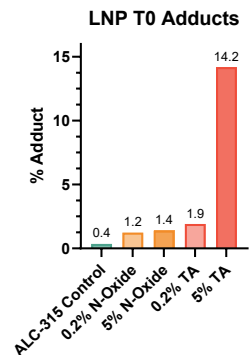
- Tail aldehyde and N-oxide content measured in 19 lots of ALC-315<sup>®</sup> and 7 lots of ALC-X, with RNA-lipid adducts measured in binary reactions
- Strong correlation between aldehyde content and RNA-lipid adduct formation

# Validating Root Cause in Raw Material

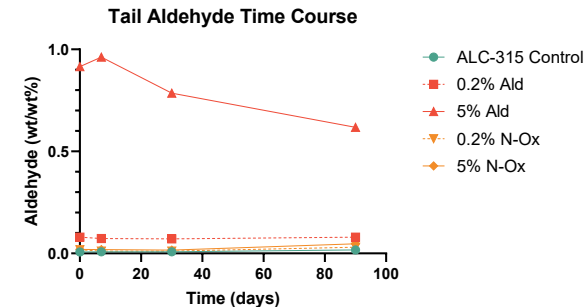
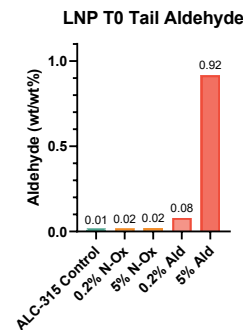


- Aldehydes are the main contributing cause of RNA-lipid adduct formation
- N-Oxide is an indirect contributor to RNA-lipid adduct formation
- In vitro **translation is primarily affected by the aldehyde** derived RNA-lipid adducts

# Validating Root Cause in LNP



- ALC-315® spiked with tail aldehyde (TA) or N-Oxide (0.2% or 5%)
- LNP formulated to monitor carryover of impurities into LNP
- LNP stored at room temperature for 3 months to monitor adduct formation
- Correlation between aldehyde content and RNA-lipid adduct formation is seen in LNP



# Impurity Impact on *in vivo* Translation (ALC-315<sup>®</sup>)

Group	Description	Storage	Measured Adducts (%)	Dose (mg/kg)	Animals
1	Control	T0 (-80°C)	0%	0.3	5F
2	Control	1 wk (RT)	1%	0.3	5F
3	Spike Low Aldehyde (2%)	T0 (-80°C)	2%	0.3	5F
4	Spike Low Aldehyde (2%)	2d (RT)	5%	0.3	5F
5	Spike Low Aldehyde (2%)	1 wk (RT)	19%	0.3	5F
6	Spike High Aldehyde (5%)	T0 (-80°C)	11%	0.3	5F
7	Spike High Aldehyde (5%)	1 wk (RT)	37%	0.3	5F
8	Control	T0 (-80°C)	0%	5.0	5F
9	Spike High Aldehyde (5%)	1 wk (RT)	37%	5.0	5F



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## • Study Parameters:

### • Pharmacodynamics

- IgG (50  $\mu$ L serum) at 24h

### • Safety/Tolerability

- Clin. Chem. (300  $\mu$ L serum) at 24h (Gr.8-9 only)
- Bodyweight at 24h
- Clinical Observation scoring at 1h, EOD, 24h

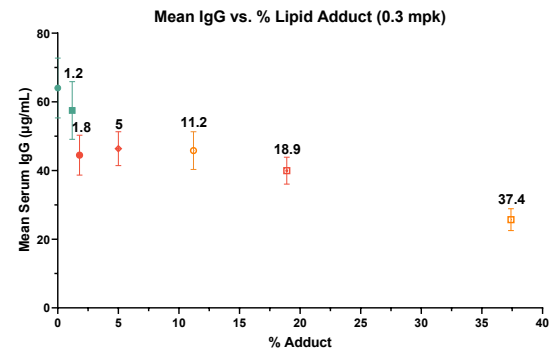
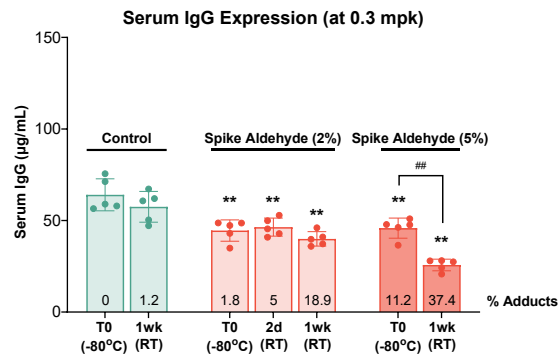
### • mRNA: CH65 IgG

- **Species:** Female CD-1, 5.5 weeks
- **Dose Administration:** i.v. bolus
- **Injection Site:** Tail vein
- **Frequency of Dosing:** Single dose
- **Monitoring Duration:** 24h
- **Biopsy/Necropsy:** 24h

# Impurity Impact on *in vivo* Translation (ALC-315<sup>®</sup>)

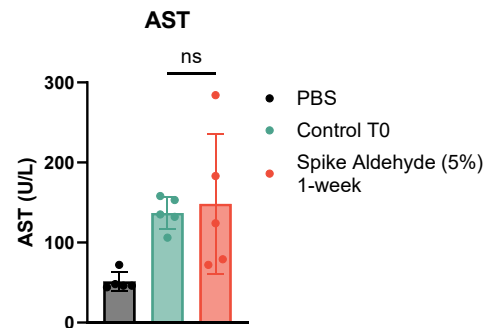
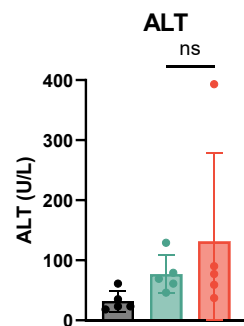
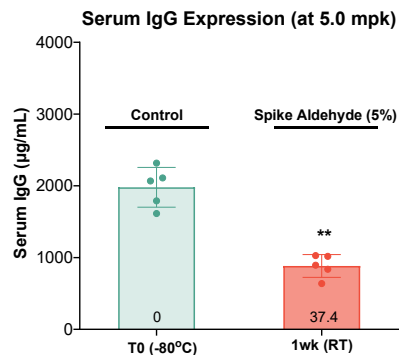
## *In vivo* Protein Expression

- Decreased serum IgG expression seen with increased % RNA-lipid adducts
- Good correlation between % RNA-lipid adducts and IgG expression ( $R^2 = 0.7824$ )



## *In vivo* Safety

- A single animal in the 5% aldehyde group had higher ALT/AST than the rest of the group
- No significant difference between ALC-315<sup>®</sup> -80°C control and 5% aldehyde group in terms of liver tolerability



# Lot Impurity Impact on *in vivo* Translation (ALC-X)

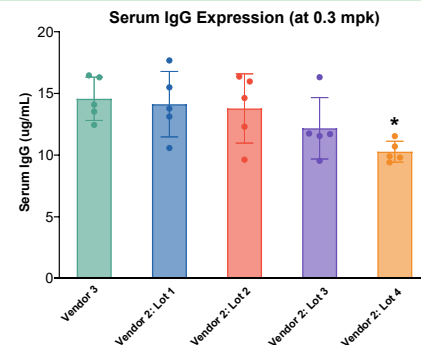
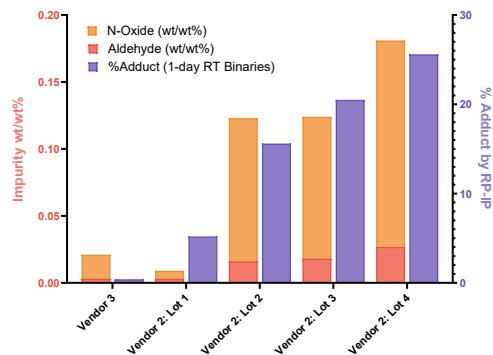
Group	Description	Dose (mg/kg)	Animals
1	Vendor 3	0.3	5F
2	Vendor 2: Lot 1	0.3	5F
3	Vendor 2: Lot 2	0.3	5F
4	Vendor 2: Lot 3	0.3	5F
5	Vendor 2: Lot 4	0.3	5F



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## *In vivo* Expression

- ALC-X lots with higher level of impurities show higher RNA-lipid adducts
- ALC-X lots showed a decreasing trend in IgG expression levels as % adduct increased
- Vendor 2: Lot 4 had the lowest activity with statistical significance



# Key Takeaways

01

## Analytical Methods

Robust analytical methods allow screening of raw materials and all LNP formulations

02

## Raw Material Screening is Crucial

Thorough screening informs on LNP long term stability and protein expression

03

## Contributing Impurities

Can guide lipid manufacturers towards directed impurity testing and new impurity acceptance criteria

04

## RNA Expression

*In vitro* and *in vivo* expression are critical to determine biological functionality of aducted RNA

# Acknowledgements

